

1 **Neuroprostheses**2 **Milos R. Popovic**3 *University of Toronto, Toronto, Ontario, Canada*4 **T. Adam Thrasher**5 *Toronto Rehabilitation Institute, Toronto, Ontario, Canada*

6

7

AQ1

8 **INTRODUCTION**

9 A neuroprosthesis, sometimes called a neural prosthesis, 48
 10 is a device that provides short bursts of electrical impulses 49
 11 to the central or peripheral nervous system to produce 50
 12 sensory and/or motor functions. Over the past four 51
 13 decades, neuroprostheses have been developed for a wide 52
 14 variety of applications. Some have achieved great success, 53
 15 such as the cochlear implants and bladder management 54
 16 stimulators that are produced in large volume worldwide. 55
 17 Other neuroprostheses, such as those for walking and 56
 18 grasping, have not yet matured to a level that creates a 57
 19 significant consumer demand. There are far too many 58
 20 neuroprostheses on the market and in development to list 59
 21 comprehensively, so this review involves the general 60
 22 features, principles, and functions of some of the most 61
 23 notable devices past and present. 62

24 **PHYSIOLOGICAL OVERVIEW**

25 In nerve cells, information is coded and transmitted as a 68
 26 series of electrical impulses called action potentials, 69
 27 which represent a brief change in cell electric potential 70
 28 of approximately 80 mV. Nerve signals are frequency 71
 29 modulated; that is, the number of action potentials that 72
 30 occur in a unit of time is proportional to the intensity of 73
 31 the transmitted signal. Typical action potential frequency
 32 is between 4 and 12 Hz. An action potential can be elicited
 33 artificially by changing the electric potential of a nerve
 34 cell or a nerve axon by inducing electrical charge into the 74
 35 cell (Fig. 1). This process, when used to produce action
 36 potentials in motor neurons to generate body function, is 75
 37 termed functional electrical stimulation (FES). 76

38 Where sufficient electrical charge is provided to a 77
 39 nerve cell, a localized depolarization of the cell wall 78
 40 occurs, resulting in an action potential that propagates 79
 41 toward the end of the axon (orthodromic propagation). 80
 42 Concurrently, an action potential will propagate backward 81
 43 towards the cell body (antidromic propagation). Typically, 82
 44 FES is concerned with orthodromic impulses, using them 83
 45 to generate muscle contractions by stimulating motor 84

nerve axons that can produce desirable body functions.
 47 Until recently, antidromic impulses were considered a
 useless side effect of FES, but there is new interest in the
 potential role of antidromic impulses in neural rehabili-
 tation.^[1]

Since generation of action potentials and their
 propagation occur in the axons, the motor nerves of the
 stimulated muscles must be intact. If peripheral axons are
 missing (if they have been cut or have degenerated, for
 example), the muscle becomes denervated and therefore
 highly resistant to electrical stimulation. However, con-
 tractions can be elicited from denervated muscles by
 applying extremely intense electrical fields across the
 muscle fibers, as demonstrated by researchers at the
 University of Vienna.^[2]

Nerves can be stimulated using monophasic or biphasic
 current or voltage pulses. The monophasic pulses are
 seldom used because they lead to unbalanced charge
 delivery to the tissues, potentially causing damage due to
 galvanic processes. Most modern FES systems implement
 biphasic current or voltage pulses, or so-called mono-
 phasic compensated pulse shapes.

Another way to activate muscles is to stimulate
 ascending axons of sensory neurons that trigger reflex
 arcs. The case where electrical stimulation is used to
 stimulate sensory neurons and thus alter reflexes or central
 nervous system functions is commonly described by the
 term neuromodulation.

25 **TECHNOLOGY**

Neuroprostheses come in many different shapes and sizes
 and serve many different purposes. The common compo-
 nents in all neuroprostheses are: 1) a power source; 2) a
 stimulus generator; 3) a user-control interface; and
 4) electrodes. Most modern neuroprostheses are batteries,
 disposable or rechargeable, as a power source. Some still
 use external AC power. Stimulus generators have been
 miniaturized dramatically over the years. Nowadays,
 commercial and laboratory-class stimulators tend to be
 lightweight (less than 1 kg) and handheld. User-control

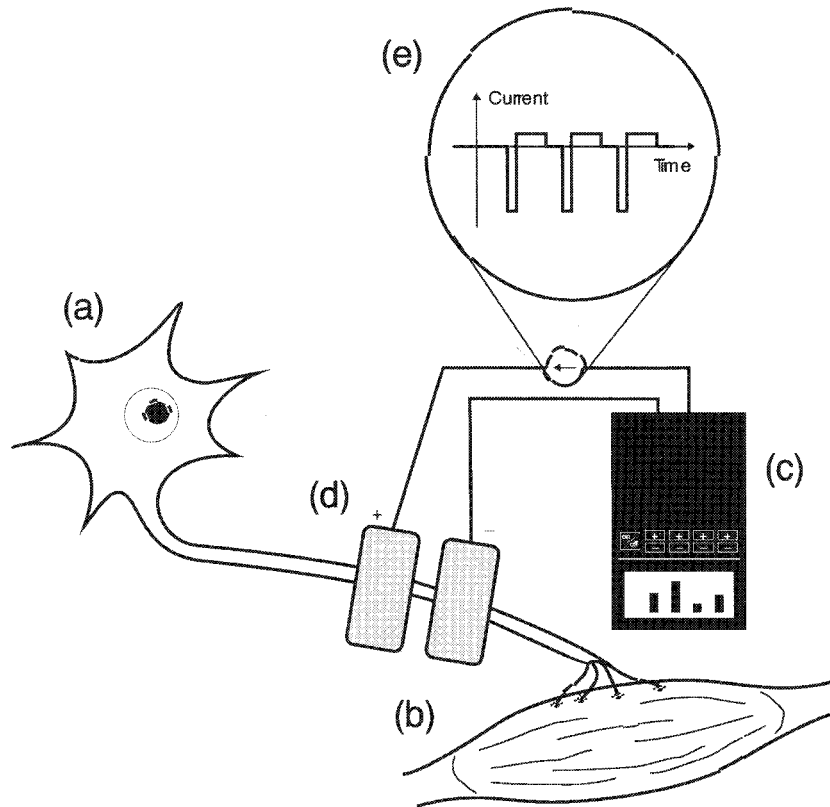


Fig. 1 Illustration of direct stimulation of a motor neuron. The cell body (a) is responsible for synthesizing input from dendrites and deciding whether or not to generate signals, which are transmitted to the corresponding muscle fibers (b). Following a stroke or spinal cord injury, muscles are impaired because motor neurons no longer receive sufficient input from the central nervous system. A neuroprosthesis (c) injects electrical current into the cell axon (d). A train of negative pulses (e) produces a series of action potentials. Depolarization occurs where negative current enters the axon at the active electrode indicated.

AQ3

85 interfaces usually consist of a simple control panel with 107
 86 standard manual controls such as switches, buttons, dials, 108
 87 and sliders, plus some kind of visual output such as light 109
 88 emitting diodes or, on more sophisticated models, a liquid 110
 89 crystal display. In addition, input devices are often 111
 90 mounted on the user's assistive devices, such as a
 91 pushbutton attached to a cane or walker. In some cases,
 92 input devices are attached to the user's clothing or body,
 93 such as an inclinometer on the shank of the leg or pressure
 94 sensors in the insole of a shoe. The most sophisticated
 95 neuroprostheses use real-time feedback control, which
 96 requires sensors such as goniometers, accelerometers, or
 97 gyroscopes to provide continuous-state feedback.

98 Nerves can be stimulated using either surface (trans-
 99 cutaneous), percutaneous, or implanted electrodes. Sur-
 100 face electrodes contact the skin (Fig. 2). They are
 101 noninvasive, easy to apply, and generally inexpensive.
 102 However, due to the impedance of the skin and the
 103 dispersion of current, much higher-intensity signals are
 104 required than with subcutaneous electrodes. Current
 105 amplitude typically ranges from 10–150 mA in surface
 106 stimulation. A major limitation is that some nerves, for

example those innervating the hip flexors, are too
 profound to be stimulated by surface electrodes. Percuta-
 neous electrodes consist of thin wires that are inserted
 through the skin and into muscular tissue, remaining in
 place for a temporary period of time. In percutaneous



Fig. 2 Reusable, self-adhesive electrodes for surface stimulation come in a variety of shapes and sizes.

AQ4

AQ5

AQ6/F2100

112 stimulation, current amplitude is rarely higher than 25 160
 113 mA. The third class of electrodes is implanted electrodes, 161
 114 which are permanently implanted. Compared to surface 162
 115 electrodes, implanted and percutaneous electrodes poten- 163
 116 tially have higher stimulation selectivity with much less 164
 117 electrical charge applied, both of which are desired char- 165
 118 acteristics of FES systems. The drawbacks are that im- 166
 119 plants require a lengthy, invasive surgical process to 167
 120 install and that percutaneous electrodes can be used only 168
 121 temporarily and may cause infection at the penetration site. 169
 122 There is a brand of miniature electrode, the BION™, 170
 123 that can be implanted via hypodermic needle.^[3] They are 171
 124 cylindrical in shape, with a diameter of 2 mm and a length 172
 125 of 15 mm. Once implanted, they are powered and 173
 126 controlled via radio waves from an external controller 174
 127 that can be worn by the patient. 175

128 HISTORICAL NOTES

129 In 46 A.D., Scribonius Largus described what may be 180
 130 considered the first neuroprosthesis: the torpedo ray, 181
 131 which is capable of generating an electric potential of 25 182
 132 to 30 V.^[4] For centuries, torpedoes were prescribed for all 183
 133 sorts of ailments, including headaches, hemorrhoids, and 184
 134 even mental illness. 185

135 Following the invention of the electrostatic generator 186
 AQ7 136 in the late 17th century, electrical discharges were found 187
 137 to excite animal preparations. Stronger discharges, and 188
 138 hence stronger biological responses, were made possible 189
 AQ8 139 when the first capacitor was invented 1745. Physicians 190
 140 began treating a wide range of diseases by applying 191
 141 electrical discharges to their patients. Benjamin Franklin 192
 142 pioneered some of these techniques. 193

143 In 1791, Luigi Galvani published his discovery that 194
 144 dissected frog legs could be stimulated by touching a 195
 145 bimetallic rod to nerve and muscle. Michael Faraday built 196
 146 the first electric generator in 1831. It introduced the 197
 147 possibility of applying a series of high-frequency elec- 198
 148 trical pulses to nerves, which is the basis for all modern 199
 149 electrical stimulation. G. B. Duchenne utilized Faradism 200
 150 extensively in the latter part of the nineteenth century to 201
 151 treat various neurological disorders. Duchenne developed 202
 152 electrodes for localizing currents, and he produced a set 203
 153 of maps of the body indicating locations called motor 204
 154 points, where electrodes can be positioned to excite 205
 155 specific muscles. 206

156 NEUROPROSTHESES FOR WALKING

157 There are many neuroprostheses that address lower- 212
 158 extremity movement. As early as 1960, Kantrowitz 213
 159 demonstrated paraplegic standing by applying continuous 214

surface FES to the quadriceps and gluteus maximus 160
 muscles of a patient with complete spinal cord injury.^[5] 161
 Around the same time, Liberson and colleagues developed 162
 a simple neuroprosthesis to correct drop foot. This 163
 common symptom in hemiplegia is characterized by a 164
 lack of dorsiflexion during the swing phase of gait, 165
 resulting in short, shuffling strides.^[6] Liberson's device, 166
 which has the distinction of being the first neuroprosthesis 167
 to receive a patent, consisted of a power supply worn on a 168
 belt, two surface electrodes positioned for stimulation of 169
 the common peroneal nerve, and a heel switch. The 170
 stimulation was activated whenever the heel lost contact 171
 with the ground, and was deactivated when the heel 172
 regained contact with the ground. 173

174 Stimulation of the common peroneal nerve causes 175
 176 contraction of the muscles responsible for dorsiflexion 176
 (i.e., tibialis anterior and extensor hallucis longus among 177
 others.). It can also trigger the flexor withdrawal reflex, 178
 which may not be desirable. The flexor withdrawal reflex 179
 occurs naturally when a sudden, painful sensation is 180
 applied to the sole of the foot. It results in flexion of the 181
 hip, knee, and ankle of the affected leg and extension of 182
 the contralateral leg in order to get the foot away from the 183
 painful stimulus as quickly as possible. To prevent this 184
 from happening during FES-assisted ambulation, Vodov- 185
 nik proposed using a low-pass filter to slow the onset of 186
 stimulation current.^[7] 187

188 Following Liberson's invention and Vodovnik's revi- 188
 189 sions, a number of drop foot stimulators were developed. 189
 Some were commercialized, for example the MikroFES 190
 (Josef Stefan Institute, Ljubljana, Slovenia) and the 191
 Odstock Dropped Foot Stimulator.^[8] The latter was 192
 shown to significantly increase walking speed and 193
 efficiency, and a carryover effect was observed in stroke 194
 patients; that is, their walking speed and efficiency 195
 without the stimulator were improved.^[9] Similar studies 196
 have reported no carryover effect.^[10] Users of the Odstock 197
 device were generally satisfied with it, but almost all of 198
 them identified the surface electrodes as problematic, and 199
 two thirds would consider an implanted system instead.^[11] 200

201 The first commercially available implanted drop foot 201
 stimulator was developed by Rancho Los Amigos Medical 202
 Centre and Medtronic Inc.^[12] The surgically implanted 203
 compounds were a radio-frequency (RF) receiver, a pulse 204
 train generator, and one bipolar electrode implanted 205
 adjacent to the peroneal nerve. An external unit worn on 206
 the belt delivered power via the RF coil and received input 207
 commands from a wireless foot switch. Despite some 208
 problems with electrode migration and infection, the 209
 device was considered successful. Since then, more 210
 reliable and easier to implant systems such as the IPPO^[13] 211
 and the Aalborg University implanted stimulator^[14] have 212
 been devised, but they are not commercially available. 213
 The latter uses input from an implanted cuff electrode 214
 around the sural nerve, which is the nerve innervating the

215 skin sensors on the sole of the foot. This system is unique 243
 216 in that it requires no external sensors. 244
 217 Most modern drop foot stimulators continue to use a 245
 218 heel switch for active input. Burrige et al. tried using 246
 219 the foot switch on the nonaffected leg, but found it was 247
 220 not preferable unless the patient was unable to reliably 248
 221 achieve heel contact on the affected leg.^[15] Vodovnik was 249
 222 one of the first to experiment with manual pushbuttons 250
 223 and EMG sensors.^[7] Other alternatives to the heel switch 251
 224 include a heel/toe switch,^[16] an array of four single-axis 252
 225 accelerometers positioned on the shank,^[17] a tilt sensor 253
 226 positioned on the shank,^[18] electroneurography,^[14] a knee 254
 227 goniometer,^[19] and the Gait Phase Detection System.^[20] 255
 228 The earliest neuroprostheses for paraplegic gait 256
 229 provided continuous stimulation to the quadriceps to 257
 230 produce a mode of gait similar to long leg-brace walking. 258
 231 Later systems used alternating bilateral quad/glut stimu- 259
 232 lation (during stance phase) out of phase with peroneal 260
 233 nerve stimulation (during swing phase). One such system 261
 234 was a six-channel stimulator developed at the University 262
 235 of Ljubljana in Slovenia.^[16] Later at the same institution, 263
 236 Kralj and colleagues described a technique for paraplegic 264
 237 gait using surface stimulation, which remains the most 265
 238 popular method today.^[21] According to Kralj's technique, 266
 239 four channels of stimulation are used. Electrodes are 267
 240 placed over the quadriceps muscles and peroneal nerves 268
 241 bilaterally. The user controls the neuroprosthesis with two 269
 242 pushbuttons attached to the left and right handles of a 270

walking frame, or on canes or crutches (Fig. 3). When the 243
 neuroprosthesis is turned on, both quadriceps are stimu- 244
 lated. The left button initiates swing phase in the left leg 245
 by briefly stopping stimulation of the left quadriceps 246
 and stimulating the peroneal nerve. This stimulation is 247
 applied suddenly so as to trigger the flexor withdrawal 248
 reflex, resulting in simultaneous hip and knee flexion as 249
 well as dorsiflexion. After a fixed period of time, peroneal 250
 nerve stimulation is stopped and quadriceps stimulation 251
 is resumed. Similarly, the right button initiates swing 252
 phase in the right leg. Kralj and colleagues successfully 253
 applied this system to more than 50 subjects with spinal 254
 cord injury. 255

Many neuroprostheses for walking have employed the 256
 basic technique described in this section. As micropro- 257
 cessor technology developed, neuroprostheses became 258
 more portable and flexible. The Parastep system uses 259
 Kralj's technique.^[22,23] It is the only neuroprosthesis for 260
 walking to receive approval from the United States Food 261
 and Drug Administration (FDA) and the first neuropros- 262
 thesis to receive FDA approval. It includes an ankle-foot 263
 orthosis to bolster ankle stiffness. The Parastep is 264
 commercially available, and more than 600 people have 265
 used it successfully. 266

A major limitation of neuroprostheses for walking 267
 that are based on surface stimulation is that the gait is 268
 slow, awkward, and unnatural looking. Perhaps a major 269
 reason for this is that the hip flexors cannot be stimulated 270

F3/AQ9

AQ10

AQ11

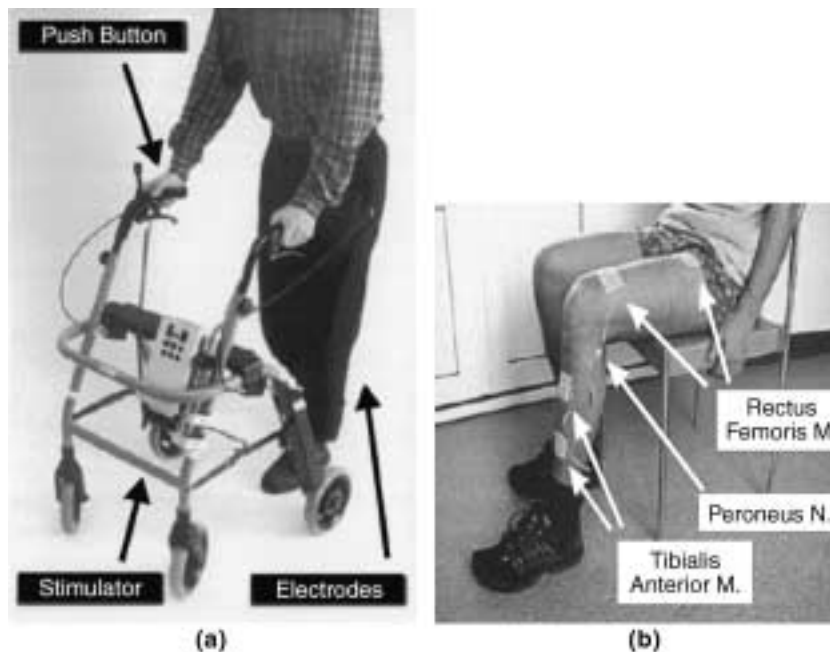


Fig. 3 The ETHZ-ParaCare walking neuroprosthesis for hemiplegic subjects and subjects with unilateral paraplegia, developed in Zurich, Switzerland. (From Ref. [38].) (a) Stimulator and push button attached to walker; (b) surface electrodes attached to legs underneath clothing.



271 directly. Therefore, hip flexion during walking must come 322
 272 from voluntary effort, which is often absent in paraplegia, 323
 273 or from the flexor withdrawal reflex (initiated by peroneal 324
 274 nerve stimulation). Implanted systems have the advantage 325
 275 of being able to stimulate the hip flexors. They also 326
 276 provide better muscle selectivity and more natural gait 327
 277 patterns. Two such systems are the Praxis24 and the 328
 278 system proposed by Kobetic, which use 24 and 32 329
 279 electrodes respectively.^[24,25] The Praxis24 system also 330
 280 enables bladder voiding. 331

281 NEUROPROSTHESES FOR REACHING 282 AND GRASPING

283 A number of neuroprostheses have been developed and 337
 284 used to assist stroke and spinal cord-injured subjects to 338
 285 improve their grasping function. The best-known grasping 339
 286 neuroprostheses are the Freehand system,^[26] the Hand- 340
 287 master NMS-1,^[27] the Bionic Glove,^[28] the NEC 341
 AQ12 288 FESMate system,^[29] the Compex Motion neuroprosthesis 342
 289 for grasping,^[30,31] and the systems developed by Rebersek 343
 290 and Vodovnik^[32] and Popovic et al.^[33] With the exception 344
 291 of the Freehand and NEC FESMate systems, all use 345
 292 surface stimulation. 346

AQ13 293 The key element for achieving the synergistic activity 347
 294 of muscles that results with reaching and grasping is the 348
 295 appropriate sequencing of electrical pulses. The available 349
 296 neuroprostheses for grasping can restore the two most 350
 297 frequently used grasping styles: the palmar and the lateral 351
 298 grasp. The palmar grasp is used to hold bigger and heavier 352
 299 objects such as cans and bottles, and the lateral grasp is 353
 300 used to hold smaller and thinner objects such as keys, 354
 301 paper, and compact discs. The lateral grasp is generated 355
 302 by first flexing the fingers to provide opposition, which is 356
 303 followed by the thumb flexion. The palmar grasp is 357
 304 generated by first forming opposition between the thumb 358
 305 and the palm, which is followed by simultaneous flexion 359
 306 of both the thumb and the fingers. 360

307 The Freehand system, manufactured and distributed by 361
 308 NeuroControl Co., U.S.A.,^[34] consists of eight implanted 362
 309 epimysial stimulation electrodes that stimulate flexion and 363
 310 extension of the fingers and the thumb in order to provide 364
 311 lateral and palmar grasp. Commands are given by an 365
 312 external position sensor that is placed on the shoulder of 366
 313 the subject's opposite arm. An additional external switch 367
 314 allows the user to choose between palmar and lateral 368
 315 grasp. This sequence is then sent via a radio frequency coil 369
 316 to the implanted unit, which generates the stimulation 370
 317 sequences for each channel. 371

318 The electrode leads are tunneled subcutaneously to the 372
 319 implanted stimulator located in the pectoral region. 373
 320 Surgical procedures to enhance both voluntary and 374
 321 stimulated hand functions are often performed in con- 375

junction with the stimulator implantation. More than 200
 quadriplegic subjects have received the Freehand neuro-
 prosthesis at more than a dozen sites around the world.
 The subjects have demonstrated the ability to grasp and
 release objects and to perform activities of daily living
 more independently when using the neuroprosthesis. The
 Freehand system is the first neuroprosthesis for grasping
 approved by the FDA. The main advantage of the
 Freehand system is that it is implanted, and the time
 needed to don and doff the system is shorter compared to
 most of the surface FES systems.

In the 1980s, the group led by Handa at Sendai
 University, Japan, developed a microcomputer-controlled
 neuroprosthesis for grasping. Soon after that, Handa's
 team proposed a system with 16 percutaneous intramus-
 cular stimulation electrodes that is both portable and
 programmable.^[29] This system consists of a NEC PC-
 98LT personal computer and an external microcontroller-
 based stimulator. The stimulator applies trapezoidal
 stimulation patterns to generate muscle contractions.
 The stimulation patterns were "cloned" from the muscle
 activity recorded during voluntary grasping movements of
 able-bodied subjects. Stimulation sequences were trig-
 gered with a pushbutton or a pneumatic pressure sensor.
 This system demonstrated that spinal cord injured subjects
 with complete C4 to C6 spinal cord lesions could reach
 and grasp. In collaboration with NEC Inc., the Sendai
 team developed a fully implantable 16-channel electric
 stimulator called the NEC FESMate. Although 200 of
 these stimulators have been manufactured,^[35] the NEC
 FESMate is not available outside of Japan.

The neuroprosthesis developed by Rebersek and
 Vodovnik was one of the first FES systems for grasp-
 ing.^[32] This neuroprosthesis has three stimulation chan-
 nels, which are used to generate grasping by stimulating
 the finger flexors and extensors and the thumb flexors.
 Although this device was developed almost three decades
 ago, it is one of the rare systems that allows a subject to
 control the stimulator via different sensory interfaces such
 as an EMG sensor, a sliding resistor, or a pressure sensor.
 This option is important because it allows the neuropros-
 thesis to be tailored to the subject. The main disadvan-
 tages of this neuroprosthesis are that donning and doffing
 times are long and that selectivity of stimulation is quite
 limited. Merelitti et al. Modified this system and used it for
 stroke subjects.^[36] They applied two channels to augment
 the elbow and fingers/wrist extension. They concluded
 that FES contributed greatly to recovery of hand and
 elbow movements in five stroke subjects, but in the
 remaining three the improvement was significant only at
 the elbow joint.

The Handmaster (Fig. 4) is a neuroprosthesis for
 grasping that is manufactured and distributed by NESS
 Ltd., located in Israel.^[27] It consists of an orthosis that has



Fig. 4 The Handmaster system manufactured and distributed by NESS Ltd., Israel. (From Ref. [27].) The flexible, adjustable unit is worn over the forearm, wrist, and palm.

409 during tenodesis grasp, significantly enhancing the
 410 strength of the grasp. Four self-adhesive surface stimu-
 411 lation electrodes provide stimulation, and the stimulator and
 412 a wrist position sensor are located on the forearm of the
 413 glove. An easy-to-use interface with three push buttons on
 414 the stimulator is used to set the stimulation parameters,
 415 and the optional audio feedback facilitates faster learning.
 416 Clinical evaluation of the Bionic Glove has indicated that
 417 it is generally beneficial to quadriplegic subjects, but only
 418 about 30% of potential users accepted it for long-term
 419 use.^[37] It is available only for clinical evaluation from the
 420 University of Alberta, Canada, and it is presently being
 421 modified into a new system called the Tetron that will
 422 provide several grasping patterns and strategies.

423 The Belgrade Grasping-Reaching System proposed by
 424 Popovic et al. is a neuroprosthesis for grasping that also
 425 provides reaching function.^[33] It consists of four stimu-
 426 lation channels, three of which are used to generate
 427 grasping function. The fourth channel is used to stimulate
 428 the triceps brachii muscle augmenting elbow extension.
 Reaching is achieved by measuring the subject's shoulder
 velocity with a goniometer and by generating a synergistic
 elbow motion by stimulating the triceps brachii muscle.
 This neuroprosthesis, similar to the system proposed by
 Rebersek and Vodovnik, requires more time to don than
 the Handmaster and is not yet commercially available.

429 The Compex Motion neuroprosthesis is a very flexible
 430 device designed to improve grasping and walking
 431 functions in both spinal cord injury and stroke patients.^[38]
 432 This multichannel surface stimulation system is program-
 433 mable and can be interfaced with any sensor system. As a
 434 four-channel neuroprosthesis for grasping, it provides
 435 both palmar and lateral grasps. It can be controlled with
 436 proportional EMG, discrete EMG, pushbuttons, or sliding
 437 resistor control strategies. Thus far, more than 50 stroke
 438 and spinal cord-injured patients have used the neuro-
 439 prosthesis in a clinical setting or at home in activities
 440 of daily living. One of the main disadvantages of this
 441 system is that it requires about eight-minutes to put on or
 442 take off.

NEUROPROSTHESES FOR BLADDER MANAGEMENT

443 Neuroprostheses have been very successful in treating
 444 lower urinary tract dysfunctions commonly associated
 445 with spinal cord injury, such as urge incontinence and
 446 urinary retention. The first attempts to electrically
 447 stimulate the bladder were made in the 1950s, when
 448 researchers sought ways to induce bladder emptying. At
 449 that time, a bladder wall stimulator was developed and
 450 implanted in three humans,^[39] and animal studies of

376 built-in flexibility to enhance and control freedom of
 377 movement within the forearm and hand, while supporting
 378 the wrist joint at a functional angle of extension. The
 379 Handmaster multiplexes a single channel of stimulation
 380 through a selected combination of surface electrodes on
 381 the inner surface of the orthosis, which effectively
 382 transforms the device into a six-channel neuroprosthesis.
 383 One stimulation channel is used to stimulate the extensor
 384 digitorum communis at the dorsal side of the forearm. The
 385 second channel stimulates the flexor digitorum super-
 386 ficialis. Electrodes are positioned over the muscles of the
 387 forearm and hand intrinsics during an initial setup session
 388 with a clinician. The setup position of the electrodes
 389 depends on the device user's specific needs. The Hand-
 390 master is controlled with an array of push buttons
 391 allowing the subject to select the operating mode and to
 392 trigger programmed movement sequences. Using the
 393 buttons, the subject can also control stimulation intensity
 394 and thumb posture, thereby adjusting the grasp to the size
 395 and the shape of the target object. Originally, the
 396 Handmaster was envisioned as a permanent orthotic
 397 system, but it is also used as an exercise and rehabilitation
 398 tool. One of the major advantages of the Handmaster is
 399 that it is easy to don and doff. It is exceptionally well
 400 designed and is one of the best neuroprostheses for
 401 grasping on the market. There are currently more than
 402 2000 in use.

403 The Bionic Glove is a neuroprosthesis designed to
 404 enhance the tenodesis grasp in subjects who have good
 405 voluntary control over wrist flexion and extension.^[28] By
 406 extending their wrist, users can cause passive finger
 407 flexion due to the limited length of the finger flexors. The
 408 Bionic Glove stimulates finger flexors and extensors

459 pelvic nerve stimulation were carried out.^[40] Later, it was 512
 460 found that electrical stimulation of the sacral anterior roots 513
 461 produces excellent results, and this led to the development
 462 of the Finetech-Brindley stimulator, which is the most
 463 widely used neuroprosthesis for bladder management 514
 464 today.^[41]

465 Attempts to manage incontinence using electrical 515
 466 stimulation began in the 1960s.^[42] It was found that 516
 467 urethral resistance could be increased by stimulating the 517
 468 muscles of the pelvic floor, vagina, and rectum using 518
 469 external electrodes.^[43] Eventually, fully implanted sys- 519
 470 tems were developed to suppress the detrusor muscle, thus 520
 471 preventing reflex incontinence and increasing bladder 521
 472 volume.^[44] Most spinal cord injuries result in reflex 522
 473 incontinence. Typically, detrusor-sphincter dyssynergia 523
 474 develops, in which the detrusor and urethral sphincter 524
 475 contract simultaneously rather than reciprocally. The 525
 476 detrusor also becomes hyper-reflexic, and the bladder 526
 477 becomes overactive. The standard treatments are anticho- 527
 478 linergic medication, which blocks the neuromuscular 528
 479 junctions, and sensory rhizotomy (surgical transection of 529
 480 the posterior sacral roots). Neuroprostheses for bladder 530
 481 management serve as a practical alternative to these 531
 482 treatments. They can also augment sensory rhizotomy. 532

483 The Finetech-Brindley stimulator has been implanted 533
 484 in more than 2000 patients, usually those who have had a 534
 485 rhizotomy.^[45] The electrodes are positioned on the 535
 486 second, third, and fourth sacral roots, bilaterally and 536
 487 extradurally. If a rhizotomy has not been performed, the 537
 488 electrodes must be implanted inside the dura to prevent 538
 489 crossover stimulation of the sensor neurons, which will 539
 490 trigger the detrusor reflex. A portable external controller 540
 491 transmits power to the implant via radio frequency coil, 541
 492 and the user initiates bladder voiding by pushing buttons 542
 493 on the external unit. Micturition is usually achieved with 543
 494 residual volumes of less than 50 mL, contributing to a 544
 495 dramatic reduction in urinary tract infections.^[46] The 545
 496 Finetech-Brindley stimulator has proven to be extremely 546
 497 robust, with only one failure expected every 80 implant- 547
 498 years.^[47] 548

499 The Medtronic Interstim stimulator is a sacral root 549
 500 implant for incontinence, using neuromodulation to 550
 501 correct the inappropriate reflex behaviour.^[48] It consists 551
 502 of fine wire electrodes inserted into the sacral foramina. 552
 503 When active, these electrodes inhibit the detrusor, but 553
 504 the mechanism of this inhibition is not yet properly 554
 505 understood. Thorough testing must be done using a 555
 506 temporary implant before permanent implantation is 556
 507 recommended. The stimulation parameters commonly 557
 508 used are a pulsewidth of 60–270 μ s and a frequency of 558
 509 10–15 Hz, with the stimulation on for 10 s, then off for 2 s. 559
 510 Current amplitude is twice the sensory threshold. The 560

AQ15 511 clinical success rate of this device is about 50%. Bladder 561

emptying has to be achieved either voluntarily or by
 means of intermittent catheterization.

COCHLEAR IMPLANTS

Cochlear implants are neuroprostheses for the hearing
 impaired who have severe (70 to 90 dB) or profound (>90
 dB) hearing loss. A long wire electrode is implanted
 directly into the cochlear duct, and electrical stimulation is
 applied to the residual spiral ganglion cells of the cochlear
 nerve. These devices were first developed in France in
 1957. Since then, cochlear implants have been refined and
 miniaturized, and now they have received widespread
 acceptance, more so than any other class of neuro-
 prostheses. More than 75,000 patients have received
 cochlear implants worldwide. Originally, few hearing-
 impaired people were eligible for cochlear implantation,
 but as the technology has improved, the selection criteria
 have expanded greatly to include a wide range of hearing
 impairments.^[49]

Due to the success and popularity of cochlear implants,
 there are many different brands on the market. Most
 brands, however, are essentially similar. Differences
 between the currently available cochlear implants mainly
 involve the number of electrode channels (12 to 22),
 speech coding strategies, and the mode of electrode stim-
 ulation.^[50] A recent study carried out at the University
 of Toronto, for example, concluded that the Clarion CI
 (Advanced Bionics, Sylmar, CA) and the Nucleus 22
 (Cochlear Corp., Sydney, Australia) cochlear implants
 were totally comparable in function or performance.^[51]
 Both devices succeeded in reducing tinnitus, thereby
 increasing word and sentence recognition, but there was
 no significant reduction in vestibular function. Among the
 implantates, 76% reported that they were satisfied with
 their implants, and 96% reported an overall positive
 impact on quality of life. Some other brands of cochlear
 implant are the COMBO 40+ system (MED-EL, Durham,
 NC), Digisonic (MXM, France), and the SOUNDTEC
 direct system (SOUNDTEC, Palo Alto, CA), most of
 which are FDA approved.

Cochlear implants generally consists of the following:
 1) an external earpiece; 2) a speech processor; and 3) an
 internally implanted unit (Fig. 5). The earpiece, usually
 very small and lightweight, is worn comfortably behind
 the ear, much like a hearing aid. It contains an ear-level or
 in-ear microphone and a radio frequency coil to transmit
 signals to the implanted components. The speech
 processor, can be in the form of a small box worn some-
 where on the body or, in some models, it is contained in
 the external unit worn behind the ear. The internal-
 ly implanted unit consists of a receiver coil located



AQ16

AQ17

F5/AQ18

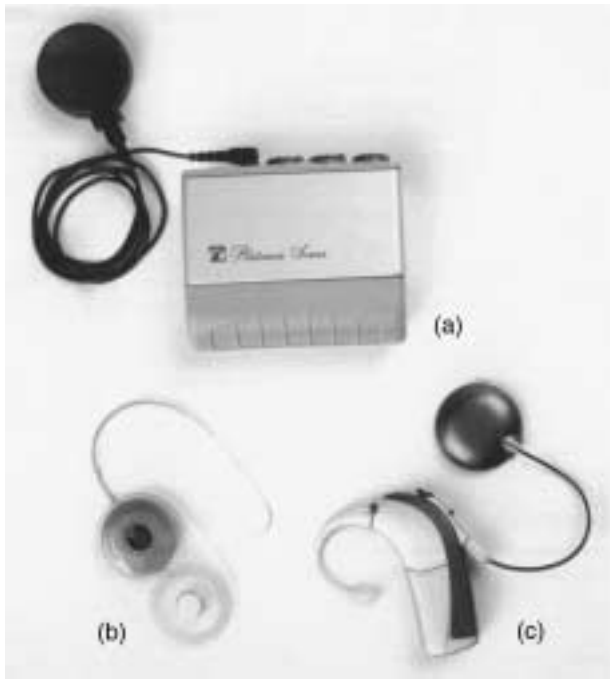


Fig. 5 The Clarion CII Bionic Ear system by Advanced Bionics (Symlar, California, USA). (a) Sound processing unit; (b) internal, implantable unit; (c) external earpiece.

584 Despite many advances and positive reports over the
 585 decades, neuroprostheses for walking and grasping have
 586 not achieved widespread approval. Most are used only for
 587 research purposes, and few have been used regularly by
 588 patients for activities of daily living. Some have been put
 589 to clinical use, and those are usually abandoned after a
 590 short period of time. The general perception among
 591 clinicians is that the neuroprostheses for grasping and
 592 walking are not fully matured and their application is
 593 often labor-intensive, whereas a favorable outcome cannot
 594 be guaranteed. Nevertheless, recent studies indicate that
 595 these tools have great potential in the rehabilitation of
 596 stroke and spinal cord-injured subjects.^[30,31,52,53] In
 597 particular, these studies indicate that a significant number
 598 of patients who were trained with these systems recover
 599 voluntary reaching, grasping, and walking functions due
 600 to intensive and repetitive training with these systems.
 601 Current efforts are focused on understanding the mech-
 602 anism of short- and long-term improvements and re-
 603 coveries observed in these patients.

604 Neuroprostheses are a new and emerging technology
 605 that has significant potential. However, implementation of
 this technology to its full potential presents numerous
 challenges that have yet to be addressed. We believe that
 the 21st century will be the century when most of these
 technical and implementation issues are resolved and
 neuroprostheses are established as one of the important
 classes of rehabilitation systems available to patients with
 disabilities ranging from spinal cord injury to blindness.

ARTICLES OF FURTHER INTEREST

REFERENCES

- 620 1. Rushton, D.N. Functional electrical stimulation and
 621 rehabilitation—An hypothesis. *Med. Eng. Phys.* **2003**, *25*
 622 (1), 75–78.
- 623 2. Reichel, M.; Breyer, T.; Mayr, W.; Rattay, F. Simulation of
 624 the three-dimensional electrical field in the course of
 625 functional electrical stimulation. *Artif. Organs* **2002**, *26*
 626 (3), 252–255.
- 627 3. Loeb, G.E. Presentation highlights: Bionic neurons
 628 (BIONs™). *J. Rehabil. Res. Dev.* **2002**, *39* (Supplement
 629 3), 5–6.
- 630 4. Schechter, D.S. Origins of electrotherapy. *N. Y. State J.*
 631 *Med.* **1971**, *71*, 997–1008.

562 underneath soft tissue in a cavity drilled in the temporal
 563 squama, and a 20–24mm insulated wire ending in a
 564 multichannel electrode array, which is inserted into the
 565 cochlear duct. Sound waves are received by the external
 566 microphone and converted into electrical signals that are
 613 input to the external speech processor. There, the signals
 568 are digitally encoded and transmitted to the internal unit
 614 via radio frequency coil. The internal unit decodes the
 615 radio signals back into elementary electrical signals to
 616 stimulate each channel of the electrode array. Therefore,
 617 the multichannel device provides a complex sound
 618 analysis similar to the physiological analysis of sound in
 573 normal patients.

CONCLUSIONS

576 The first modern FES devices were developed over 40
 577 years ago, and since then there has been a great deal of
 578 innovation resulting in scores of new neuroprostheses.
 579 The most successful of these technologies, in terms of
 580 consumer acceptance, are cochlear implants—more than
 581 30,000 units have been implanted worldwide. Bladder
 582 management stimulators have also achieved wide accept-
 583 ance, but not quite to the same degree.

AQ20

- 632 5. Kantrowitz, A. A Report of the Maimonides Hospital. In 691
633 *Electronic Physiologic Aids*; Brooklyn: New York, 1960. 692
- 634 6. Liberson, W.T.; et al. Functional electrotherapy: Stimu- 693
635 lation of the peroneal nerve synchronized with the swing 694
636 phase of the gait of hemiplegic patients. *Arch. Phys. Med.* 695
637 *Rehabil.* **1961**, *42*, 101–105. 696
- 638 7. Vodovnik, L.; Long, C.; Reswick, J.B.; Lippay, A.; 697
639 Starbuck, D. Myo-electric control of paralyzed muscles. 698
640 *IEEE Trans. Biomed. Eng.* **1965**, *12* (3), 169–172. 699
- 641 8. Taylor, P.N.; Burrige, J.H.; Dunkerley, A.L.; Wood, D.E.; 700
642 Norton, J.A.; Singleton, C.; Swain, I.D. Clinical use of the 701
643 Odstock Dropped Foot Stimulator: Its effect on speed and 702
644 effort of walking. *Arch. Phys. Med. Rehabil.* **1999**, *80* (12), 703
645 1577–1583. 704
- 646 9. Taylor, P.N.; Burrige, J.H.; Dunkerley, A.L.; Lamb, A.; 705
647 Wood, D.E.; Norton, J.A.; Swain, I.D. Patients' percep- 706
648 tions of the Odstock Dropped Foot Stimulator. *Clin.* 707
649 *Rehabil.* **1999**, *13* (5), 439–446. 708
- 650 10. Granat, M.H.; Maxwell, D.J.; Ferguson, A.C.; Lees, K.R.; 709
651 Barbenel, J.C. Peroneal stimulator; evaluation for the 710
652 correction of spastic drop foot in hemiplegia. *Arch. Phys.* 711
653 *Med. Rehabil.* **1996**, *77* (1), 19–24. 712
- 654 11. Taylor, P.N.; Mann, G.; Wood, D.E.; Hobby, J. In *Pilot* 713
655 *Study to Evaluate the Safety and Efficacy of an Implanted* 714
656 *Dropped Foot Stimulator*, From Technology to Market: 715
657 Bridging the Gap, Proceedings of the International 716
658 Functional Electrical Stimulation Society, Maroochydore, 717
659 Australia, July 1–5, 2003. 718
- 660 12. Waters, R.L.; McNeal, D.; Perry, J. Experimental cor- 719
661 rection of foot-drop by electrical stimulation of the per- 720
662 oneal nerve. *J. Bone Jt. Surg., Am.* **1975**, *57-A*, 1047– 721
663 1054. 722
- 664 13. Strojnik, P.; Acimovic, R.; Vavken, E.; Simic, V.; Stanic, 723
665 U. Treatment of drop foot using an implantable peroneal 724
666 underknee stimulator. *Scand. J. Rehab. Med.* **1987**, *19*, 725
667 37–43. 726
- 668 14. Haugland, M.K.; Sinkjaer, T. Cutaneous whole nerve 727
669 recordings used for correction for footdrop in hemiplegic 728
670 man. *IEEE Trans. Biomed. Eng.* **1995**, *3*, 307–317. 729
- 671 15. Burrige, J.H.; Taylor, P.N.; Hagan, S.A.; Wood, D.E.; 730
672 Swain, I.D. The effects of common peroneal stimulation on 731
673 the effort and speed of walking: A randomized controlled 732
674 trial with chronic hemiplegic patients. *Clin. Rehabil.* **1997**, 733
675 *11* (3), 201–210. 734
- 676 16. Strojnik, P.; Kralj, A.; Ursic, I. Programmed six-channel 735
677 electrical stimulator for complex stimulation of leg 736
678 muscles during walking. *IEEE Trans. Biomed. Eng.* 737
679 **1979**, *26*, 112–116. 738
- 680 17. Willemsen, A.T.M.; Bloemhof, F.; Boom, H.B.K. Auto- 739
681 matic stance-swing phase detection from accelerometer 740
682 data for peroneal nerve stimulation. *IEEE Trans. Biomed.* 741
683 *Eng.* **1990**, *37*, 1201–1208. 742
- 684 18. Dai, R.; Stein, R.B.; Andrews, B.J.; James, K.B.; Wieler, 743
685 M. Application of tilt sensors in functional electrical 744
686 stimulation. *IEEE Trans. Rehabil. Eng.* **1996**, *4* (2), 63–72. 745
- 687 19. Sweeney, P.C.; Lyons, G.M. Fuzzy gait event detection in 746
688 a finite state controlled FES drop foot correction system. *J.* 747
689 *Bone Jt. Surg., Br.* **1999**, *81-B* (Suppl. I), 93. 748
- 690 20. Pappas, I.P.; Popovic, M.R.; Keller, T.; Dietz, V.; Morari, 749
M. A reliable gait phase detection system. *IEEE Trans.*
Neural Syst. Rehabil. Eng. **2001**, *9* (2), 113–125.
21. Kralj, A.; Bajd, T.; Turk, R. Enhancement of gait
restoration in spinal injured patients by functional elec-
trical stimulation. *Clin. Orthop. Relat. Res.* **1988**, *223*, 34–
43.
22. Graupe, D.; Davis, R.; Kordylewski, H.; Kohn, K.H.
Ambulation by traumatic T4-12 paraplegics using func-
tional neuromuscular stimulation. *Crit. Rev. Neurosurg.*
1998, *8*, 221–231.
23. Graupe, D.; Kohn, K.H. Functional neuromuscular stim-
ulator for short-distance ambulation by certain thoracic-
level spinal-cord-injured paraplegics. *Surg. Neurol.* **1998**,
50, 202–207.
24. Davis, R.; Houdayer, T.; Andrews, B.J.; Barriskill, A. In
Paraplegia: Implanted Praxis24±FES System for Multi-
Functional Restoration, Proceedings of the International
Functional Electrical Stimulation Society, Sendai, Japan,
1999.
25. Kobetic, R.; Triolo, R.; Morsolais, E. Muscle selection and
walking performance of multichannel FES systems for
ambulation in paraplegia. *IEEE Trans. Rehabil. Eng.* **1997**,
5, 23–28.
26. Smith, B.; Peckham, P.H.; Keith, M.; Roscoe, D. An
externally powered, multichannel, implantable stimulator
for versatile control of paralyzed muscle. *IEEE Trans.*
Biomech. Eng. **1987**, *34* (7), 499–508.
27. Ijzerman, M.; Stoffers, T.; 't Groen, F.; Klatte, M.; Snoek,
G.; Vorsteveld, J.; Nathan, R.; Hermens, H. The NESS
handmaster orthosis: Restoration of hand function in C5
and stroke patients by means of electrical stimulation. *J.*
Rehabil. Sci. **1996**, *9* (3), 86–89.
28. Prochazka, A.; Gauthier, M.; Wieler, M.; Kenwell, Z. The
Bionic Glove: An electrical stimulator garment that
provides controlled grasp and hand opening in quadriple-
gia. *Arch. Phys. Med. Rehabil.* **1997**, *78*, 1–7.
29. Hoshimiya, N.; Handa, Y. A master-slave type multichan-
nel functional electrical stimulation (FES) system for the
control of the paralyzed upper extremities. *Automedica*
1989, *11*, 209–220.
30. Popovic, M.R.; Hajek, V.; Takaki, J.; Bulten, A.;
Zivanovic, V. In *Restoration of Reaching and Grasping*
Functions in Hemiplegic Patients with Severe Arm
Paralysis, From Technology to Market: Bridging the
Gap, Proceedings of the International Functional Electrical
Stimulation Society, Maroochydore, Australia, July 1–5,
2003.
31. Adams, M.; Takes, V.; Popovic, M.R.; Bulten, A.;
Zivanovic, V. In *Restoration of Grasping Functions in*
Patients with Quadriplegia, From Technology to Market:
Bridging the Gap, Proceedings of the International
Functional Electrical Stimulation Society, Maroochydore,
Australia, July 1–5, 2003.
32. Rebersek, S.; Vodovnik, L. Proportionally controlled
functional electrical stimulation of hand. *Arch. Phys.*
Med. Rehabil. **1973**, *54*, 168–172.
33. Popovic, D.; Popovic, M.; Stojanovic, A.; Pjanovic, A.;
Radosavljevic, S.; Vulovic, D. In *Clinical Evaluation of*
the Belgrade Grasping System, Proceedings of the 6th



- 750 Vienna International Workshop of FES, Vienna, Austria, 790
751 1998. 791
- 752 34. Smith, B.; Peckham, P.; Keith, M.; Roscoe, D. An 792
753 externally powered, multichannel, implantable stimulator 793
754 for versatile control of paralyzed muscle. IEEE Trans. 794
755 Biomech. Eng. **1987**, *34*, 499–508. 795
- 756 35. Takahashi, K.; Hoshimiya, N.; Matsuki, H.; Handa, Y. 796
757 Externally powered implantable FES system. Jpn. J. Med. 797
758 Electron. Biol. Eng. **1999**, *37*, 43–51. 798
- 759 36. Merletti, R.; Acimović, R.; Grobelnik, S.; Cvilak, G. 799
760 Electrophysiological orthosis for the upper extremity in 800
761 hemiplegia: Feasibility study. Arch. Phys. Med. Rehabil. 801
762 **1975**, *56* (12), 507–513. 802
- 763 37. Popovic, D.; Stojanovic, A.; Pjanovic, A.; Radosavljevic, 803
764 S.; Popovic, M.; Jovic, S.; Vulovic, D. Clinical evaluation 804
765 of the Bionic Glove. Arch. Phys. Med. Rehabil. **1999**, *80* 805
766 (3), 299–304. 806
- 767 38. Popovic, M.R.; Keller, T.; Pappas, I.P.I.; Dietz, V.; Morari, 807
768 M. Surface-stimulation technology for grasping and 808
769 walking neuroprostheses. IEEE Trans. Eng. Med. Biol. 809
770 Mag. **2001**, *20*, 82–93. 810
- 771 39. Boyce, W.H.; Lathem, J.E.; Hunt, L.D. Research related to 811
772 the development of an artificial electrical stimulator for the 812
773 paralyzed human bladder: A review. J. Urol. **1964**, *91*, 41– 813
774 51. 814
- 775 40. Bors, E.; Comarr, E. *Neurological Urology: Physiology of* 815
776 *Micturition, Its Neurological Disorders and Sequelae*; 816
777 Karger Verlag, 1971. 817
- 778 41. Brindley, G.S.; Polkey, C.E.; Rushton, D.N.; Cardozo, L. 818
779 Sacral anterior root stimulators for bladder control in 819
780 paraplegia: The first 50 cases. J. Neurol. Neurosurg. 820
781 Psychiatry **1986**, *49*, 1104–1114. 821
- 782 42. Caldwell, K.P.S. The electrical control of sphincter in- 822
783 continence (preliminary communication). Lancet **1963**, *2*, 823
784 174–175. 824
- 785 43. Alexander, S. Research and clinical experience in the 825
786 treatment of the neurogenic bladder by electronic implant 826
787 and prosthesis. Paraplegia **1968**, *6* (3), 183–193. 827
- 788 44. Rijkhoff, N.J.M.; Wijkstra, H.; van Kerrebroeck, P.E.V.; 828
789 Debruyne, F.M.J. Urinary bladder control by electrical 829
stimulation: Review of electrical stimulation techniques
in spinal cord injury. NeuroUrol. Urodyn. **1997**, *16*, 39–
53.
45. Schurch, B.; Rodic, B.; Jeanmonod, D. Posterior sacral
rhizotomy and intradural anterior sacral root stimulation
for treatment of the spastic bladder in spinal cord injured
patients. J. Urol. **1997**, *157*, 610–614.
46. Creasey, G.H. Restoration of bladder, bowel, and sexual
function. Top Spinal Cord Inj., Rehabil. **1999**, *5* (1), 21–
32.
47. Vastenholt, J.M.; Ijzerman, M.J.; Buschman, H.P.J.;
Snoek, G.J.; van der Aa, H.E. In *Seven Year Follow-Up
of Sacral Anterior Root Stimulation and Sacral Posterior
Root Rhizotomy for Bladder Control in Patients with a
Spinal Cord Injury, Complications and Quality of Life*,
Proceedings of the International Functional Electrical
Stimulation Society, Cleveland, Ohio, 2001.
48. Siegel, S.W. Management of voiding dysfunction with an
implantable neuroprosthesis. Urol. Clin. North Am. **1992**,
19 (1), 163–170.
49. Lenarz, T. Cochlear implants, selection criteria and
shifting borders. Acta-Rhino-Laryngol. Belg. **1998**, *52*
(3), 183–199.
50. Gstoettner, W.; Adunka, O.; Hamzavi, J.; Baumgartner,
W.D. Rehabilitation of hearing-impaired patients with
cochlear implants, a review. Wien. Klin. Wochenschr.
2000, *112* (11), 464–472.
51. Higgins, K.M.; Chen, J.M.; Nedzelski, J.M.; Shipp, D.B.;
McIlmoyl, L.D. A matched-pair comparison of two
cochlear implant systems. J. Otolaryngol. **2002**, *31* (2),
97–105.
52. Popovic, M.B.; Popovic, D.B.; Sinkjaer, T.; Stefanovic, A.;
Schwirtlich, L. Restitution of reaching and grasping
promoted by functional electrical therapy. Artif. Organs
2002, *26* (3), 271–275.
53. Thrasher, T.A.; Popovic, M.R. In *FES-Assisted Walking
for Rehabilitation of Incomplete Spinal Cord Injury*, From
Technology to Market: Bridging the Gap, Proceedings of
the International Functional Electrical Stimulation Society,
Maroochydore, Australia, July 1–5, 2003.